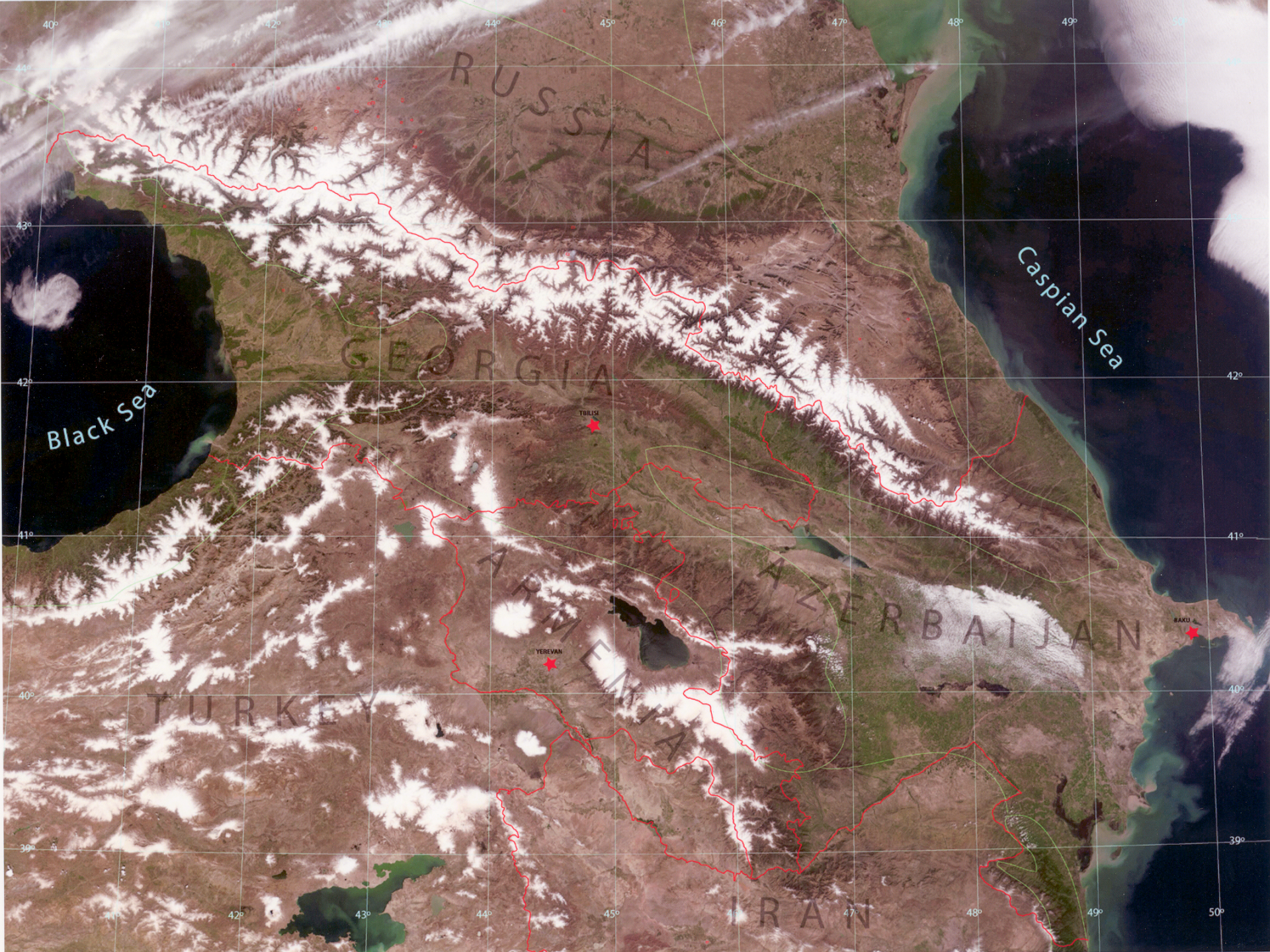


# **Biotechnological Developments in Georgia**

**Giorgi Kvesitadze**

A decorative graphic consisting of several sets of concentric circles in a lighter shade of blue, located in the bottom right corner of the slide.



RUSSIA

GEORGIA

ARMENIA

AZERBAIJAN

TURKEY

IRAN

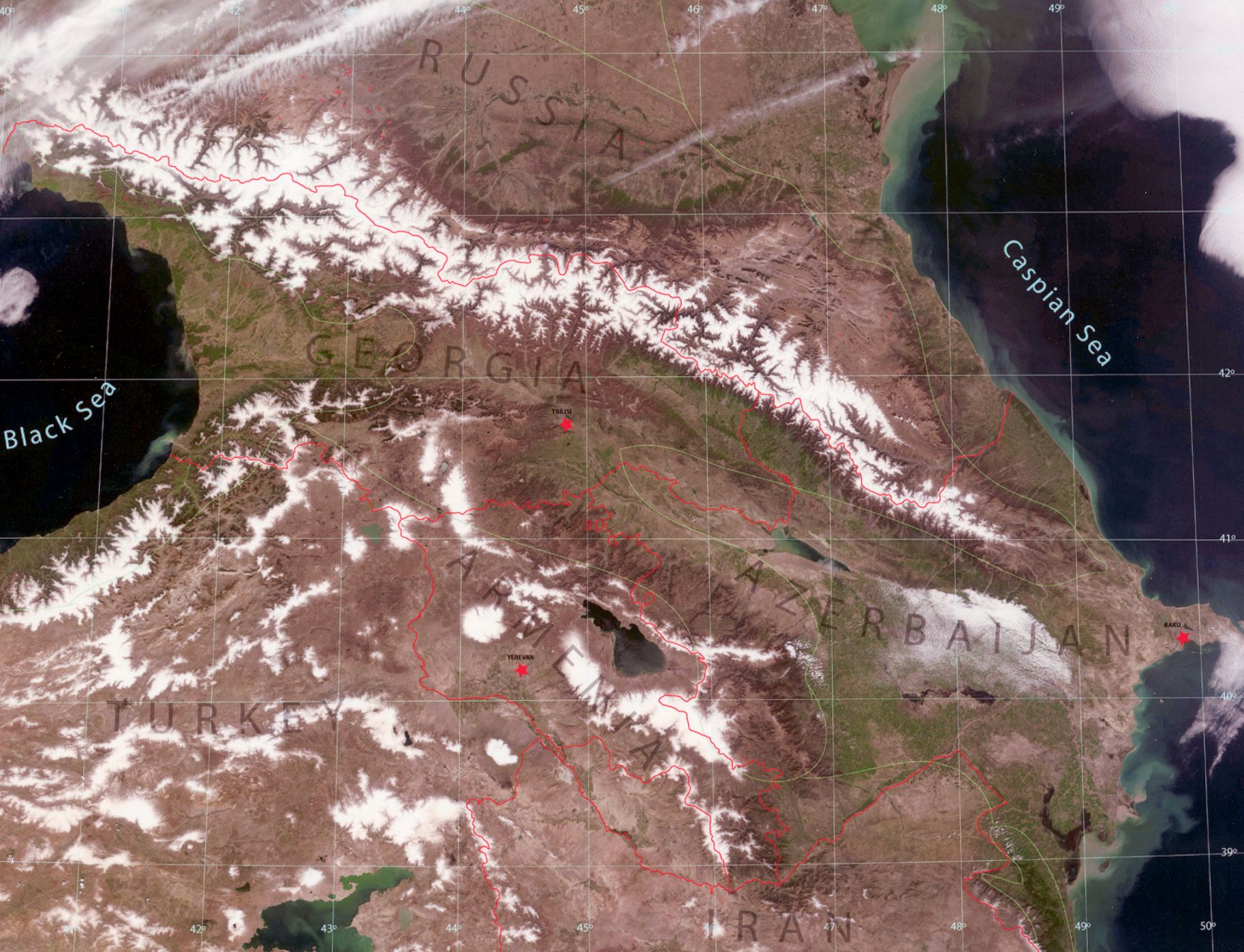
Black Sea

Caspian Sea

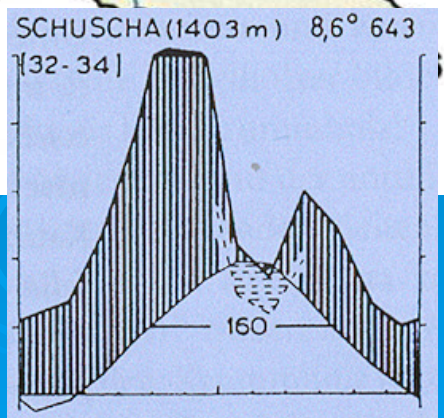
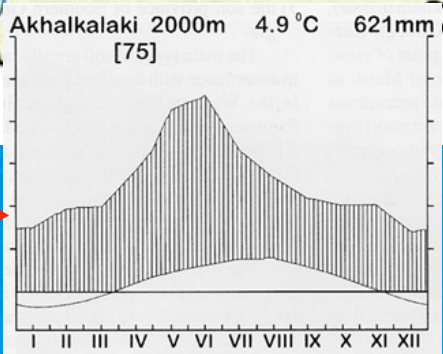
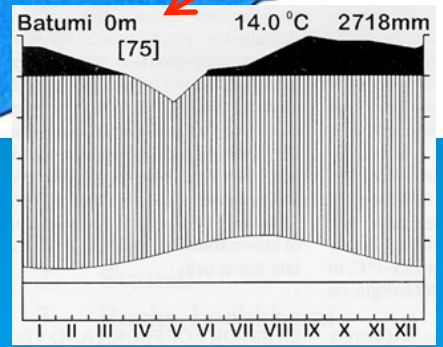
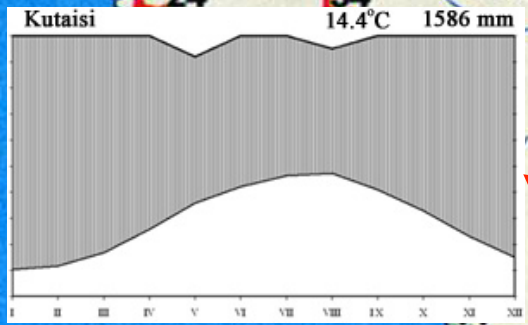
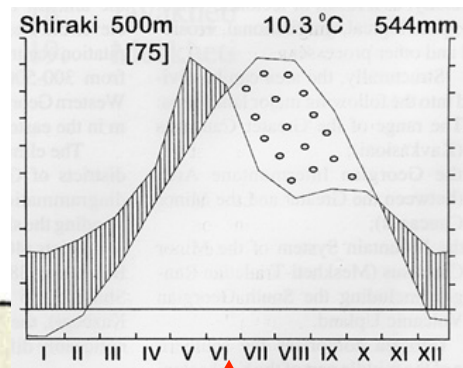
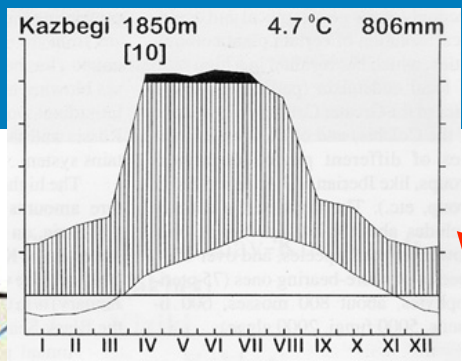
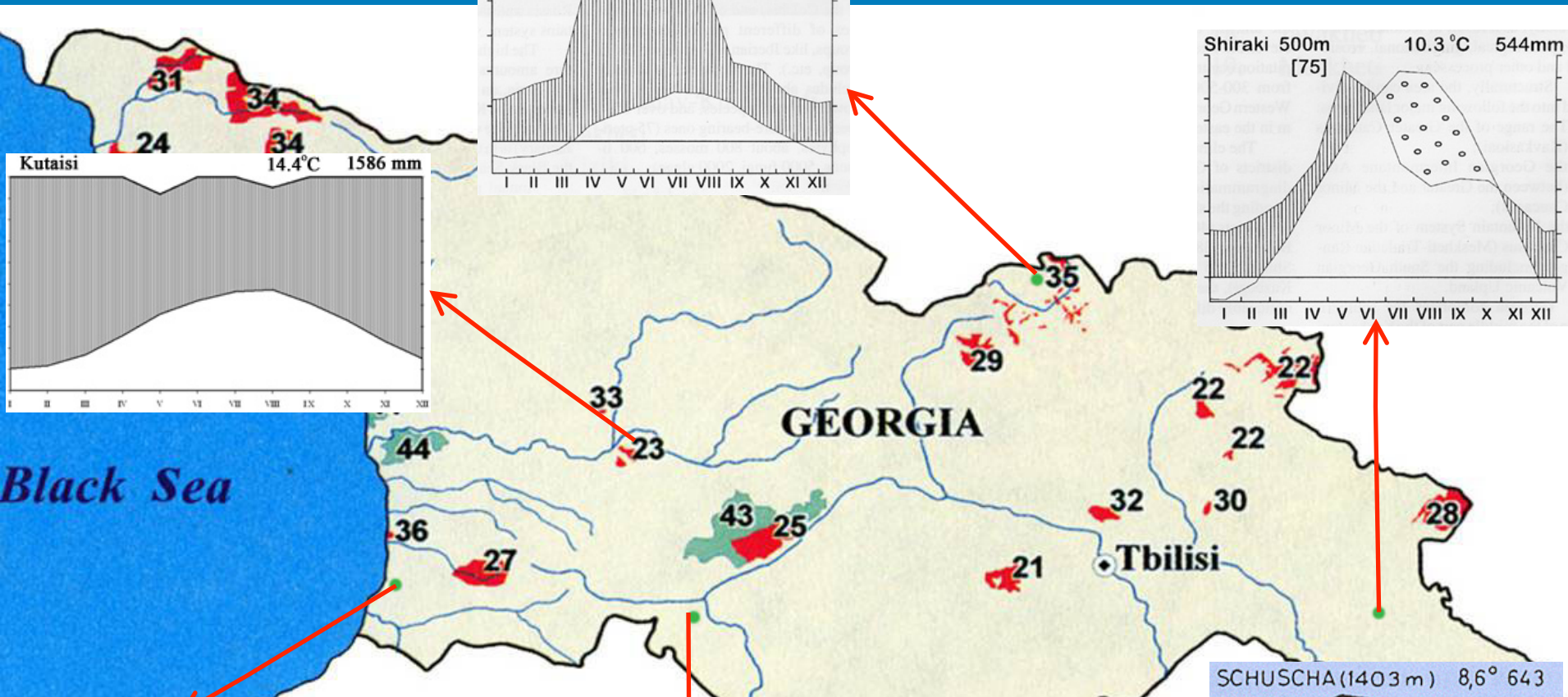
TBILISI

YEREVAN

BAKU



69.700 km<sup>2</sup>



# Black Sea

- 1 Mountain meadow soddy, soddy peat and primitive soils
- 2 Mountain chernozems and soddy mountain meadow soils of Southern foothills
- 3 Brown soil
- 4 Forest cinnamonic soils
- 5 Soddy-calcareous soils
- 6 Leached and calcareous mountain chernozems
- 7 Chernozems
- 8 Grey cinnamonic soil, with natric soil complexes
- 9 Red and yellow soils
- 10 Podzolic and grey podzols

- 11 Raw humus sulphatic, solonch and natric soils
- 12 Boddy soils
- 13 Alluvial calcareous and uncalcareous soils
- 14 Meadow calcareous soils of Eastern Georgia
- 15 Different types of primitive soils



# Plants taxonomic diversity of the Caucasus

Caucasus                      6350 species  
   1100 endemics

Georgia                      5100 species  
   1000 endemics

Alps                              4500 species  
   350 endemics

Above 100 flavonoids of different structure (catechins, proantocyanidines, flavones, flavonols, oxycinnamic acids) have been isolated from the wild and cultural endemic flora of Georgia (*Citrus*, *Rhododendron*, *Vitis*, *Laurocerasus*, *Vaccinium*).

Among tasted plants more than 20 industrially important endemic sources of flavanoids are revealed.

# Caucasian plants medicinal preparations

## **RHODOPESUM**

*Rhododendron ungerii* Trautv.

Flavonoids (catechins, proanthocyanidins).

## **FATSIFLOGINUM**

*Fatsia japonica* (Thunb.) Decne et Panache.

Triterphenyl glucoside.

## **FLARONIN**

*Astragalus falcatus* Lam.

Flavonoid glucoside, robinin.

## **TSARUBOL**

*Paliurus spina-christi* Mill.

Neutral lipids, phospholipids, flavonoids, including proanthocyanidins.

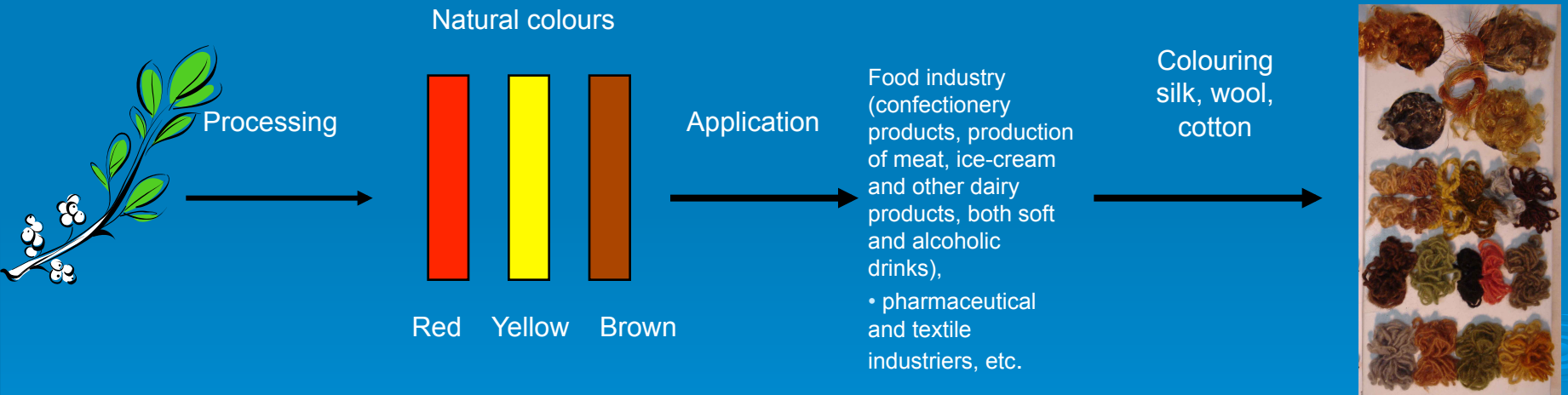
## **COLCHICINE**

*Colchicum speciosum*

Colchicin-(S)-N-(5,6,7,9, tetrahydro-1,2,3,10-tetramethoxy-9-oxobenzo-[ ]-heptalene-7-yl)

# Plant Food Colours

From the diverse Caucasus flora the technology of the production of red, yellow and brown colours has been worked out. These colours contain harmless natural pigments and polyphenols similar to green tea, particularly (-) epigallocatechingallate and other biologically active compounds. It has been established that plants polyphenols almost completely abolish the influence of harmful radioactive isotope – Strontium-90 by absorbing and eliminating it out of the human organism before reaching marrow. These compounds possess antioxidant and antibactericide activities. They block the development of different tumours and cancer diseases, prevent tooth caries, promote digestion, decrease cholesterol level in blood, regulate blood pressure. The use of these colours as products, possessing antiradiation and anticarcinogenic properties is allowed by the Pharmacological Committee of Georgia.



**Production and use of natural colours derived from plants**



# Bacteria

The collections of microorganisms of different taxonomic groups: bacteria, actinomycetes, mycelial fungi and yeasts at Dumishidze Institute of Biochemistry and Biotechnology (DIBB), have been created on the base of strains isolated from different soil climatic zones of Georgia, and account above 5000 strains, including extremophiles. Among the collections have been identified strains active producers of enzymes, secondary metabolites (vitamins, antibiotics, organic acids etc.), medicinal polysacchrides and actively degrading organic contaminants of different structure.



*Bacillus* sp. C-7 - active producer of proteolytic enzymes in deep conditions; alkaline protease

*Rhodococcus* sp. BKM Ac 1170, mutant strain 44 - producers of secondary metabolites: carotinoides, B group vitamins

DIBB collection of bacteria isolated from different soil climatic zones of Georgia, since 1970. Above 3500 strains



*Rhodococcus* strains degrading naphthalene



*Mycobacterium* sp. 235. *Rhodococcus* sp. 227 - degraders of TNT, *Pseudomonas* sp 44, *Rhodococcus* sp. 74, mineral and crude oil.

# ACTINOMYCETES

Production of enzymes, antibiotics, vitamins, hormones, etc.

→ Strains active producers of proteolytic enzymes: (*Streptomyces* sp. 28, halophil, *Streptomyces* sp. 154, halophil. Chitinase (*Streptomyces* sp. 101; *Streptomyces* sp. 105).



DIBB collection of *Actinomycetes*, including extremophiles, isolated from different soil-climatic zones of Georgia. Since 1965. Above 2000 strains.

Degradation of organic contaminants.

→ Strains degrading oil hydrocarbons (open chain and cyclic, including PAH):  
*Streptomyces* sp 138, halophil;  
*Streptomyces* sp 166, alkaliphil;  
*Streptomyces* sp.118.  
3,4-benzpyrene - *Streptomyces* sp. 51, alkaliphil, *Streptomyces* sp. 69, alkaliphil;  
Naphthalene-*Streptomyces* sp. 6, alkaliphil, etc.



# MYCELIAL FUNGI



DIBB collection of Mycelial fungi strains (basidial and microscopic), including extremophiles, isolated from different soil-climatic zones of Georgia. Since 1967. Above 4000 strains.

Producers of enzymes *Aspergillus oryzae* 27, *Aspergillus awamori* 11, *Aspergillus niger* 47, *Aspergillus batatae* 21, *Aspergillus usami* 15, ( $\alpha$ -amylase, glucoamylase); *Mucor* sp. (acid protease); *Aspergillus tamari* T 53; *Aspergillus wentii* 52, *Aspergillus versicolor* 83, *Absidia* sp. K-61, *Chaetomium* sp. S-77, *Pleurotus ostreatus* 88, *Lentinus edodes* 1016, *Pycnoporus coccineus* 310; *Trametes versicolor* 13; *Daedalea quereina* 350; *Phlebia radiata* 312 (cellulases, xylanases); *Pseudotamella gilbosa* 22; *Trametes versicolor* 5; *Pleurotus eringii* 45; *Pleurotus ostreatus* 108, *Cerrena maxima* 681; *Funalia trogii* 146; *Lentinus edodes* 779 (Mn-peroxidase, laccase), etc.

Producers of medicinal polysaccharides: *Ganoderma lucidum* 447; *Lentinus edodes* 123, *Pleurotus ostreatus* 24, *Coriolus hirsutus* 68, *Phellinus robustus* 29, etc.

Several dozens of mycelial fungi strains degrading wide spectrum of organic contaminants. *Fusarium* sp. 26; *Mucor* sp. 1; *Trichotecium* sp. 65; *Aspergillus niger* 35; *Penicillium* sp. 51, *Trichoderma viride* 9, etc.

# EDIBLE MUSHROOMS

More than 7000 species of basidiomycetous fungi are widespread in various ecosystems of Georgia: semi-desert and forests, alpine and subtropics, mountains and steppes. Besides the local species there are strangers from Mediterranean areas, the European Plane, the Iranian Highland and Central Asia. 570 species, 62 varieties and forms of *Agaricales* basidiomycetes are described (1). The most species-rich genera are *Russula*, *Lactarius*, *Mycena*, *Inocybe*, *Clitocybe*, *Coprinus*, *Marasmius*, *Collybia*, *Pholiota*, *Agaricus*, *Pluteus*, *Polyporus*, *Hygrophorus*, *Lepiota*, *Oudemansiella*, *Calocybe*, *Omphalina*, *Panaeolus*, *Pleurotus*, *Agrocybe*, *Stropharia*. In addition to these fungi, 258 species of *Aphylophorales* basidiomycetes have been described (2). Analysis showed several ecological groups including mycorrhizal (182 species), xylotrophs (128), litter saprotrophs (132), humus saprotrophs (91), coprotrophs (13), herbotrophs (7), bryotrophs (5), sphagnetrophs (4), carbotrophs (4) and mycotrophs (4).

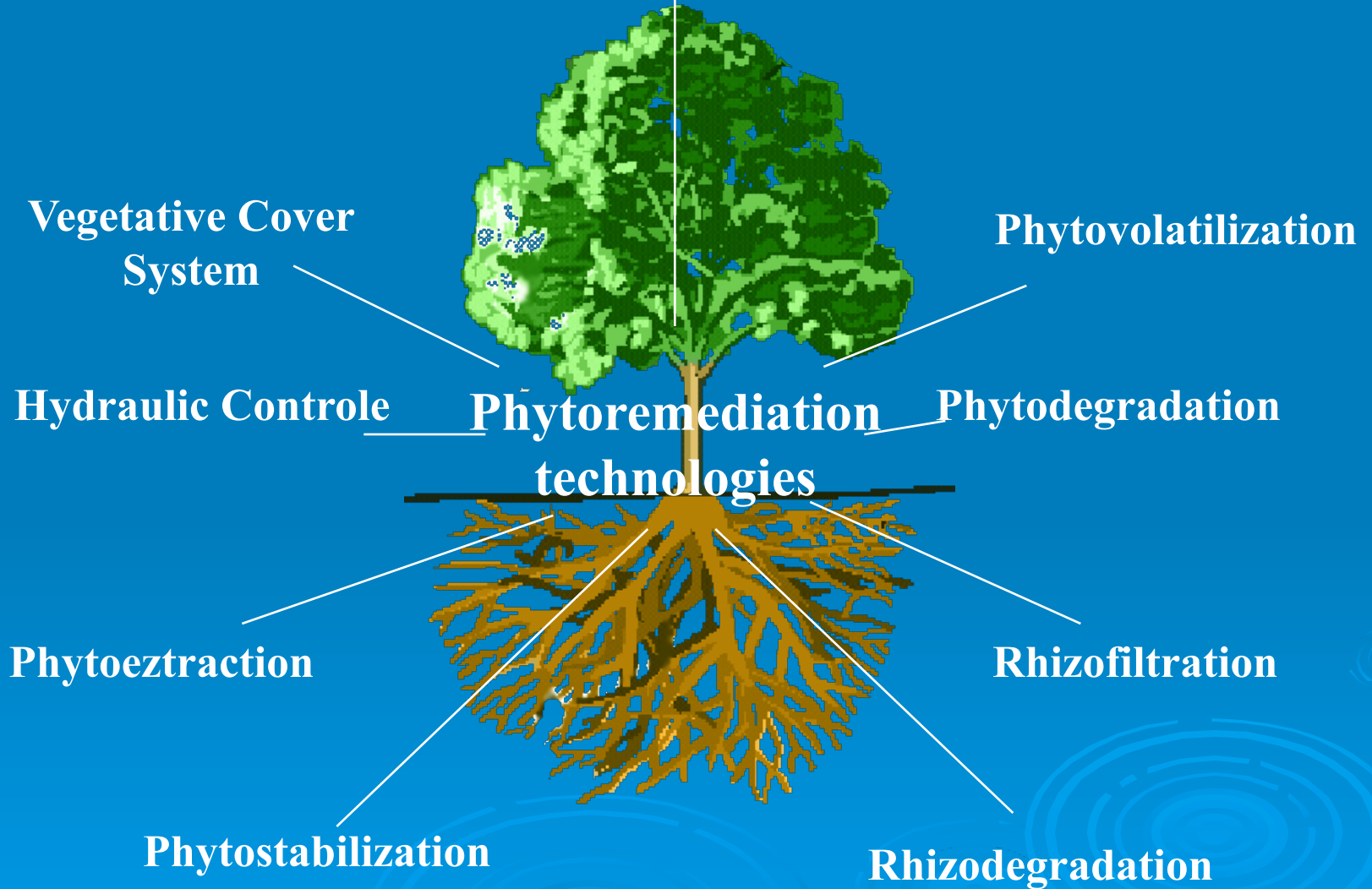
Especially widespread and widely used in Western Georgia is *Amanita caesarea*.

1. Nakhutsrishvili V., (1975). Agarics mushrooms of Georgia, Tbilisi, 209 pages
2. Flora of Cryptogrammic Plants and Fungi of Georgia., (1996). Tbilisi, 885 pages

# Bacteriophage application in the Infection Pathology

- **Intesti** Bacteriophage-in intestinal infections:  
*Shigella (Flexner, Zonne, Newcastle),*  
*Salmonella (Paratyphus A, Paratyphus B,*  
*Typhimurium, Enteritidis, Choleraesuis, Oranienburg),*  
enteropathogenic serotypes of *E.coli* (10 types),  
*Proteus, Enterococcus, Staphylococcus,*  
*Pseudomonas.*
- **Pyobacteriophage**-against suppurative infections  
(*Staphylococcus, Streptococcus, E.coli, Pseudomonas,*  
*Proteus*).

**“Green filter”**



At Durmishidze Institute of Biochemistry and Biotechnology, since 1960 the investigations to characterize higher plants potential to absorb and metabolize xenobiotics have been carried. The evaluation of over 100 plants potential to degrade above 150 contaminants of different structure: alkanes, alkenes, aromatic including polycyclic hydrocarbons, phenols, aldehydes, etc. has been established. As a result of over 20 years investigation the concept of "Green filter" has been elaborated, envisaging the purposeful planting of selected plants around or along the sources of contamination (highways, chemical and metallurgical factories, gas and oil pipelines, etc).

## Green Filter Technology

Plants can very effectively hinder the emission of environmental contaminants. Good examples are exhaust gases of motorcars: carbon monoxide, nitrogen and sulfur oxides, aromatic hydrocarbons, etc.

A “Green filter” 30m wide of five layers (four lines of shrubs, mulberry and fustic, 70 cm high, and a fifth line of maple, birch, and elm 6-8 m high, plant age 10-20 years) planted along a highway reduces carbon monoxide concentrations in the air by 60-70%.



## Plants according to their ability to assimilate benzene and toluene from air

Group of plants	Amount of absorbed aromatic hydrocarbon by 1 kg of fresh leaves, for 24 hours, mg	P l a n t s	
Strong absorbers	1.0 – 10.0	Maple ( <i>Acer campestre</i> ) Oleaster ( <i>Elaeagnus angustifolia</i> ) Locust ( <i>Robina pseudoacacia</i> ) Wild pear ( <i>Pyrus caucasica</i> ) Walnut ( <i>Juglans regia</i> ) Almond-tree ( <i>Amigdalus communis</i> ) Cherry-tree ( <i>Cerasus avium</i> ) Amorpha ( <i>Amorpha fruticosa</i> ) Cherry-tree ( <i>Cerasus vulgaris</i> ) Chestnut ( <i>Castanea sativa</i> )	Apple-tree ( <i>Malus domestica</i> ) Zelkova ( <i>Zelcova caprinifolia</i> ) Poplar ( <i>Populus canadensis</i> ) Ryegrass ( <i>Lolium perene</i> ) Lilac ( <i>Siringa vulgaris</i> ) Weeping willow ( <i>Salix</i> ) Catalpa ( <i>Catalpa bignonioides</i> ) Platan-tree ( <i>Platanus orientalis</i> ) Sophora ( <i>Sophora japonica</i> )
Average absorbers	0.1 – 1.0	Alder ( <i>Alnus barbata</i> ) Asp ( <i>Populus tremula</i> ) Elm ( <i>Ulmus filicea</i> ) Ash ( <i>Fraximus excelsior</i> ) Tea ( <i>Camellia sinensis</i> L.) Persimmon ( <i>Diospyros lotus</i> ) Bay laurel ( <i>Laurus nobilis</i> )	Gleditdchia ( <i>Gledt schia triacanthos</i> ) Kidney ( <i>Phaseolus vulgaris</i> ) Pine ( <i>Pinus</i> ) Pine ( <i>Pinus eldarica</i> ) Thuja ( <i>Tuja</i> ) Apricot ( <i>Prunus armenicana</i> ) Vine ( <i>Vitis vinifera</i> )
Weak absorbers	0.001 – 0.1	Fir ( <i>Picea abies</i> ) Mulberry ( <i>Morus alba</i> ) Lime-tree ( <i>Tilia cauxasica</i> ) Reed ( <i>Phragmites communis</i> ) Maize ( <i>Zea mays</i> ) Wild plum ( <i>Prunus divaricata</i> ) Kiwi ( <i>Apteryx australis</i> ) Aloe ( <i>Aloe</i> ) Medlar ( <i>Mespilus germanica</i> ) Rose ( <i>Rosa</i> ) Platan-tree ( <i>Platanus</i> )	Cypress ( <i>Cupressus sempervirens</i> var. <i>Pyramidalis</i> ) Geranium ( <i>Pelargonium roseum</i> ) Privet ( <i>Ligustrum vulgare</i> ) Fig ( <i>Ficus carica</i> ) Pomegranate ( <i>Punica granatum</i> ) Rhododendron ( <i>Rhododendron ponticum</i> ) Peach-tree ( <i>Persica vulgaris</i> ) Potato ( <i>Solanum tuberosum</i> ) Tomato ( <i>Lycoperssicum esculentum</i> ) Pussy-willow ( <i>Salix alba</i> ) Cherry-plum ( <i>Prunus yeshupaltii</i> )

George Kvesitadze, Academy of Sciences of Georgia, Tbilisi;  
Gia Khatisashvili, Academy of Sciences of Georgia, Tbilisi;  
Tinatin Sadunishvili, Academy of Sciences of Georgia, Tbilisi;  
Jeremy Ramsden, Inst. of Advanced Study, Collegium  
Basilea, Basel, Switzerland

## Biochemical Mechanisms of Detoxification in Higher Plants

### Basis of Phytoremediation

Plants play a key role in purifying the biosphere of the toxic effects of industrial activity. This book shows how systematic application of the results of investigations into the metabolism of xenobiotics (foreign, often toxic substances) in plants could make a vastly increased contribution to planetary well-being. Deep physiological knowledge gained from an accumulation of experimental data enables the great differences between the detoxifying abilities of different plants for compounds of different chemical nature to be optimally exploited. Hence planting could be far more systematically adapted to actual environmental needs than is actually the case at present. The book could form the basis of specialist courses in universities and polytechnics devoted to environmental management, and advanced courses in plant physiology and biochemistry, for botany and integrative biology students. Fundamental plant physiology and biochemistry from the molecular level to whole plants and ecosystems are interwoven in a powerful and natural way, making this a unique contribution to the field.

**Contents:** Plant and environment.- The fate of pollutants in plant cell.- Ecotechnologies based on phytoremediation

G. Kvesitadze  
G. Khatisashvili  
T. Sadunishvili  
J. Ramsden

## Biochemical Mechanisms of Detoxification in Higher Plants

Basis of Phytoremediation

 Springer

# Biotechnologies developed at Durmishidze Institute of Biochemistry and Biotechnology

- **Production of 20 different oxidative and hydrolytic enzymes isolated from microorganisms and plants (amylases, cellulases, proteases, xylanases, Mn-peroxidase, laccase, etc)**
- **Pectin (from 10 varieties of endemic plants)**
- **Carotenoides (from bacteria)**
- **Bacteriophages, the biological effective means against plant pathogenic bacteria**
- **Edible and medicinal mushrooms;**
- **Medicinal polysaccharides**

# Organizations involved in biotechnology developments:

**Durmishidze Institute of Biochemistry and Biotechnology;**  
**Georgian Technical University;**  
**Institute of Molecular Biology;**  
**Eliava Institute of Bacteriophage, Microbiology and Virology;**  
**Institute of Botany;**  
**Institute of Zoology;**  
**Institute of Subtropical Cultures;**  
**Georgian Agrarian University.**  
**Tbilisi State University**

*Above 25 average and small companies participate in Georgia in production and distribution of biotechnological products.*

# Central Caucasus



# KAZBEGI



# Svaneti. Ushguli.



# Shatili





# Tusheti

